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J. Nat. Prod., 1993, 56 (11), 2016-2018• DOI: 10.1021/np50101a027 • Publication Date (Web): 01 July 2004

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BIEMNASTEROL, A NEW CYTOTOXIC STEROL WITH THE RARE 22,25-DIENE SIDE CHAIN, ISOLATED FROM THE MARINE SPONGE *BIEMNA* SP.

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ABSTRACT.—Biemnasterol [2], a new sterol with a 22,25-diene side chain possessing cytotoxic activity, has been isolated from the Okinawan marine sponge *Biemna* sp. and the structure elucidated on the basis of spectroscopic data and chemical means.

A variety of unconventional sterols with unusual side chains and nuclei have been isolated from marine sponges (1-3). Among these was the sterol 24β methylcholesta-5,7,22,25-tetraen-3B-ol [1], first isolated from a Hawaiian sponge Ciocalypta sp. as a major component of a sterol mixture (4). It contained the $\Delta^{5,7}$ sterol nucleus and interesting 22,25-diene side chain and provided strong evidence for the proposed intermediacy of a 22,25-diene in the biosynthesis of sterols (5). During our investigations on bioactive substances from Okinawan marine organisms (6-8), we recently examined extracts of the Okinawan sponge Biemna sp. and have isolated a new sterol, biemnasterol [2], possessing the rare 22,25-diene side chain, together with a known compound 1. In this paper we describe the isolation and structure elucidation of compound 2; its structure was confirmed by the chemical transformation of compound 1 into 2.

The sponge *Biemna* sp. was collected at Unten Harbor, Okinawa Island and kept frozen until used. The MeOH extract was partitioned between EtOAc and 1M NaCl aqueous solution. The EtOAc-soluble fraction was subjected to Si gel flash cc eluted with hexane/EtOAc and CHCl₃/MeOH, followed by purification by a Sephadex LH-20 column to give biemnasterol [**2**](0.0004%, wet wt) along with compound **1** (0.0015%).

Biemnasterol [2] was obtained as a colorless needle; the molecular formula was indicated to be $C_{28}H_{44}O_3$ by hrfabms $(m/z \ 429.3346 \ [M+H]^+, \Delta -2.3 \ mmu)$

and contained seven degrees of unsaturation. The ¹³C-nmr spectrum showed the resonances attributed to six olefinic carbons, two sp³ oxymethines, one oxygenated sp³ quaternary carbon, seven sp³ methylenes, five sp^3 methines, two sp^3 quaternary carbons, and five methyls. These signals corresponded well to those observed in the ¹³C spectrum of compound 1 except for the presence of an additional sp³ oxymethine and an oxygenated sp³ quaternary carbon and the absence of two olefinic carbons. The positions of the two oxygenated carbons were deduced as C-5 and C-6 on the basis of ¹H-¹H COSY cross peaks for H-6/H-7 and HMBC correlations for H-6/C-5, H-6/C-7, H-6/C-8, H-7/C-5, H-7/C-9, and H-7/C-14. From these observations, biemnasterol [2] was inferred to be the 5,6-dihydroxy derivative of 1. The structure of biemnasterol [2], including the stereochemistry of the hydroxyl groups, was firmly established by chemical transformation of compound 1 into 2 according to a literature method (Scheme 1) (9). Treatment of the acetate of 1 with Na₂Cr₂O₂ afforded an α -hydroxyl ketone 3, which was reduced with NaBH₄ to give a 1,2-diol 4 as a major product. Deprotection of the 3-acetoxyl group of 4furnished the triol 2. The eims, ¹H-nmr, and ir spectral data as well as the optical rotation of compound 2 thus prepared from 1 were identical with those of compound 2 of the natural specimen. The structure of biemnasterol was, therefore, concluded to be 24β-methylcholesta-7,22,25-tirene-3 β ,5 α ,6 β -triol [2].



SCHEME 1. a: Ac_2O /pyridine, room temperature, 24 h. b: $Na_2Cr_2O_7 \cdot 2H_2O/Ac_2O/HOAc/C_6H_6$, 0°, 4h. c: $NaBH_4$ /THF/iPrOH, room temperature, 48 h. d: 0.5 M KOH/MeOH, room temperature, 14 h.

Biemnasterol [2] exhibited cytotoxicity against murine lymphoma L1210 and human epidermoid carcinoma KB cells in vitro, with IC₅₀ values of 3.0 and 1.3 μ g/ml, respectively.

EXPERIMENTAL

GENERAL METHODS.—Optical rotations were recorded on a JASCO DIP-4 digital polarimeter. Uv and ir spectra were taken on JASCO Ubest-35 and JASCO Report-100 spectrometers, respectively. ¹H- and ¹³C-nmr spectra were recorded on JEOL JMN GX-270 and EX-400 spectrometers. Eims and fabms were obtained on a JEOL JMS DX-303 and a JEOL HX-110 spectrometer, respectively. Wako C-300 Si gel was used for cc, and tlc was carried out on Merck Si gel GF₂₃₄.

SPONGE MATERIAL.—The sponge *Biemna* sp. (Order Poecilosclerida; Family Desmacellidae; Gray, 1867), collected by netting at Unten Harbor, Okinawa Island, was kept frozen until used. The specimen was a very dark brown to purple black sponge when preserved, with some foreign material in the mesohyl. Skeleton a loose unispicular or bispicular reticulation of styles without fibre development. Numerous large sigmas throughout the mesohyl. Styles 552–612×12–13 μ m; sigmas 96 μ m, rephides 210 μ m long. The voucher specimen (SS-857) was deposited at the Faculty of Pharmaceutical Sciences, Hokkaido University.

ISOLATION.—The MeOH extract of the sponge (1.7 kg, wet wt) was evaporated under reduced pressure, and the residue (36 g) was partitioned between EtOAc (400 ml \times 3) and 1M NaCl (400 ml). A portion (432 mg) of the EtOAc-

soluble material (877 mg) was subjected to Si gel flash cc (50×3 cm) with gradient elution of EtOAc in hexane (0–100%) and MeOH in CHCl₃ (50– 100%). The fraction (29 mg) eluted with 20% EtOAc in hexane was further purified by Sephadex LH-20 column (50% MeOH in CHCl₃; 120×2.5 cm) to give compound 1 (23 mg). The fraction (71 mg) eluted with 50% MeOH in CHCl₃ was separated by the second Si gel column (30×1.5 cm) eluted with 0–100% MeOH in CHCl₃. The fraction (9 mg) eluted with 10% MeOH in CHCl₃ was subjected to a Sephadex LH-20 column (50% MeOH in CHCl₃; 120×2.5 cm) to give compound 2 (3.1 mg).

Compound 1.—¹H nm (CDCl₃) $\delta_{\rm H}$ 5.58 (1H, dd, J=5.5 and 2.6 Hz, H-6), 5.39 (1H, dd, J=5.5 and 2.6 Hz, H-7), 5.22-5.27 (2H, m, H-22 and H-23), 4.68–4.72 (2H, m, H₂-26), 3.62 (1H, m, H-3), 2.72(1H, m, H-24), 2.45(1H, ddd, J=8.0, 4.4, and 2.0 Hz, H₄-4), 2.27 (1H, m, H₅-4), 2.08 (2H, m, H-20 and H,-12), 1.97 (1H, m, H-9), 1.86-1.92 (3H, m, H-14, H_a-1, and H_a-2), 1.75 (1H, m, H_a-16), 1.73(1H, m, H₄-15), 1.67(3H, s, H₃-27), 1.62(2H, m, H_2-11 , 1.59 (1H, m, H₄-2), 1.25-1.36 (5H, m, H_b-1, H_b-12, H_b-15, H_b-16, and H-17), 1.08 (3H, d, J=6.6 Hz, H₃-28), 1.03 (3H, d, J=6.6 Hz, H₃-21), 0.94 (3H, s, H_3 -19), 0.63 (3H, s, H_3 -18); ¹³C nmr (CDCl₃) δ_{c} 38.4 (t, C-1), 32.0 (t, C-2), 70.4 (d, C-3), 40.8 (t, C-4), 141.3 (s, C-5), 119.6 (d, C-6), 116.3 (d, C-7), 139.8 (s, C-8), 46.3 (d, C-9), 37.0 (s, C-10), 21.1 (t, C-11), 39.1 (t, C-12), 42.9 (s, C-13), 54.5 (d, C-14), 23.5 (t, C-15), 28.3 (t, C-16), 55.7 (d, C-17), 12.0 (q, C-18), 16.3 (q, C-19), 40.2 (d, C-20), 20.9 (q, C-21), 131.6 (d, C-22), 135.7 (d, C-23), 43.6 (d, C-24), 149.8 (s, C-25), 108.8 (t, C-26), 20.6 (q, C-27), 18.9 (q, C-28); eims m/z [M]⁺ 394 (100), 361 (75), 335 (40), 271 (25), 253 (35),

Biemnasterol [2].-Colorless solid: mp 241- $242^{\circ}; \{\alpha\}^{19}D - 7.6^{\circ} (c = 0.43, MeOH); uv (MeOH)$ λ max 204 (ε 12700); ir (KBr) ν max 3400, 2950 cm^{-1} ; ¹H nmr (CD₃OD) δ_{H} 5.30 (1H, m, H-7), 5.25 (2H, m, H-22 and H-23), 4.70 (2H, m, H₂-26), 3.96 (1H, m, H-3), 3.53 (1H, m, H-6), 2.71 (1H, m, H-24), 1.66 (3H, s, H₃-27), 1.08 (3H, d, J=7.0 Hz, H₃-21), 1.05 (3H, s, H₃-19), 1.03 (3H, $d_J = 6.6 \text{ Hz}, H_3 - 28), 0.64 (3H, s, H_3 - 18); {}^{13}C \text{ nmr}$ $(CDCl_3) \delta_c 30.8 (t, C-1), 32.0 (t, C-2), 67.7 (d, C-2)$ 3), 39.2 (t, C-4), 76.0 (s, C-5), 73.7 (d, C-6), 117.6 (d, C-7), 143.9 (s, C-8), 43.8 (d, C-9), 37.1 (s, C-10), 22.9 (t, C-11), 39.5 (t, C-12), 43.5 (s, C-13), 54.7 (d, C-14), 22.0 (t, C-15), 27.8 (t, C-16), 56.0 (d, C-17), 12.3 (q, C-18), 18.8 (q, C-19), 40.2 (d, C-20), 20.9 (q, C-21), 135.5 (d, C-22), 131.8 (d, C-23), 43.6 (d, C-24), 149.8 (s, C-25), 108.8 (t, C-26), 20.6 (q, C-27), 18.9 (q, C-28); eims m/z (rel. int. %) $[M-H_2O]^{+}$ 410 (50), $[M-2H_2O]^{+}$ 392 (30), 377 (35), 287 (10), 269 (25), 251 (30), 123 (85), 43 (100); hrfabms m/z 429.3346 (calcd for $C_{28}H_{45}O_{3}[M+H]^{+}$ 429.3369).

3B-Acetoxy-5a-bydroxy-24B-methylcholesta-7,22,25-trien-6-one [3].—Compound 1 (8.1 mg) was acetylated with $Ac_2O(0.5 \text{ ml})$ and pyridine (2 ml). After evaporation of the solvent, the crude product was dissolved in HOAc (1 ml), Ac₂O (2 ml), and C_6H_6 (1 ml), to which $Na_2Cr_2O_7 \cdot 2H_2O$ (6.2 mg) was added at 0° and stirred for 4 h at 0° . After addition of $H_2O(2 \text{ ml})$, the reaction mixture was extracted with EtOAc and the organic phase was purified by a Si gel column [20×1.5 cm; hexane-EtOAc (5:1)] to give **3** (4.3 mg): $[\alpha]^{19}$ D -5.0° (c=0.10, ClCH₂CH₂Cl); ir (KBr) ν max 3400, 2900, 1720, 1680, 1250 cm⁻¹; ¹H nmr $(CDCl_3)\delta_{H}$ 5.65 (1H, m, H-7), 5.24–5.28 (2H, m, H-22 and H-23), 5.10(1H, m, H-3), 4.70(2H, m, H,-26), 2.72 (1H, m H-24), 2.03 (3H, s, Ac), 1.68 $(3H, s, H_3-27), 1.08(3H, d, J=6.9 Hz), 1.03(3H,$ $d_{J} = 6.6 \text{ Hz}$, 0.96 (3H, s, H₃-19), 0.60 (3H, s, H₃-18); eims m/z (rel. int. %) [M]⁺ 468 (8), $[M-HOAc]^+$ 408 (6), 390 (20), 374 (18), 251 (20), 123 (90), 43 (100).

 3β -Acetoxy-24 β -methylcholesta-7,22,25-triene-5 α ,6 β -diol [4].—NaBH₄ (1.5 mg) was added to the solution of compound **3** (3.4 mg) in iPrOH (1 ml), and the mixture was stirred at room temperature for 48 h. Then H₂O (1 ml) was added, and the reaction mixture was extracted with EtOAc. The organic phase was purified by a Si gel column [20×1.0 cm; hexane-EtOAc (2:1)] to give **4** (1.1 mg) (10): [α]²¹D +40° (c=0.11, CHCl₃); ir (KBr) ν max 3450, 2950, 1720, 1270 cm⁻¹; ¹H nmr (CDCl₃) $\delta_{\rm H}$ 5.32–5.35 (1H, m, H-7), 5.24–5.27 (2H, m, H-22 and H-23), 5.15 (1H, m, H-3), 4.70 (2H, m, H₂-26), 3.61 (1H, m, H-6), 2.71 (1H, m, H-24), 2.03 (3H, s, Ac), 1.68 (3H, s, H₃-27), 1.09 (3H, s, H₃-19), 1.08 (3H, d, J=6.6 Hz), 1.02 (3H, d, J=6.6 Hz), 0.60 (3H, s, H₃-18); eims m/z (rel. int. %)[M-H₂O]⁺ 452 (38),[M-H₂O-HOAc]⁺ 392 (100), 377 (43), 363 (55), 269 (60), 251 (25), 123 (95).

24 β -Methylcholesta-7,22,25-triene-3 β ,5 α ,6 β triol [2].—Compound 4(1.1 mg) was treated with 0.5 M KOH/MeOH (1.5 ml) for 14 h at room temperature. After usual workup, compound 2 (0.9 mg) was obtained, whose ir, ¹H-nmr, eims, and [α]D data were all identical with those of a natural specimen of biemnasterol [2].

ACKNOWLEDGMENTS

We thank Mr. Z. Nagahama for his help with sponge collection, Prof. T. Sasaki (Kanazawa University) for cytotoxicity test, and Dr. J. Fromont, James Cook University of North Queensland, for identification of the sponge. This work was partly supported by a Grant-in-Aid from Toray Science Foundation and a Grant-in-Aid for Scientific Research from the Ministry of Education, Science, and Culture of Japan.

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Received 14 April 1993